HELINI™
Purefast Shrimp Nucleic acid Mini spin prep Kit

[Optimized for Shrimp biological samples]

Cat. No. 2010- 25 Preps
Cat. No: 2010 - 50 Preps
Cat. No: 2010 - 100 Preps

Handbook

www.helini.in
info@helini.in
helinibiomolecules@gmail.com

www.helini.in

Kit components

No. of reactions Catalogue Number	25 2010	50 2010	100 2010
Tissue lysis buffer	20ml	40ml	80ml
Binding Buffer	10ml	20ml	40ml
Elution Buffer	14ml	28ml	60ml
Wash Buffer-1	15ml	30ml	60ml
Wash Buffer-2	12ml	24ml	48ml
Spin columns with collection tube	25	50	100
Collection tubes	75	150	300
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Storage

Kits packed in two boxes. One carton box contains Proteinase K that should be stored in -20C. Another Box contains remaining consumables that has to be store in room temperature.

Technical Assistance

For technical assistance and more information, please contact;
0091-44-24490433
helinibiomolecules@gmail.com

Reagent Preparation

Add the indicated volume of ethanol (96-100%) to Wash Buffer I (concentrated) and Wash Buffer II (concentrated) prior to first use:

	Cat.No:2010 – 25 prep	
	Wash buffer-1	**Wash Buffer-2**
Concentrated Buffer	15ml	12ml
Ethanol [96 – 100%] to add	11ml	48ml
Total volume	**26ml**	**60ml**

	Cat.No:2010 – 50 prep	
	Wash buffer-1	**Wash Buffer-2**
Concentrated Buffer	30ml	24ml
Ethanol [96 – 100%] to add	22ml	96ml
Total volume	**42ml**	**120ml**

	Cat.No:2010 – 100 prep	
	Wash buffer-1	**Wash Buffer-2**
Concentrated Buffer	60ml	48ml
Ethanol [96 – 100%] to add	44ml	192ml
Total volume	**104ml**	**240ml**

Safety Information

When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. Discard sample and assay waste according to your local safety regulations.

Introduction

HELINI Purefast Nucleic acid Mini spin prep Kit is designed for rapid and cost-effective small-scale preparation of high-quality nucleic acid from tissue and biological samples. The kit utilizes an exclusive silica-based membrane technology in the form of a convenient spin column. The standard procedure takes less than 30 minutes following cell lysis and yields purified viral nucleic acid. Isolated Viral nucleic acid can be used directly in RT-PCR/PCR.

Principle

Cells are lysed during a short incubation with chaotropic salt, which immediately inactivates all nucleases. Cellular nucleic acids bind selectively to special glass fibres pre-packed in the purification filter tube. Bound nucleic acids are purified in a series of rapid "wash and spin" steps to remove contaminating cellular components. Finally low salt elution releases the Nucleic acids from the glass fibre. This simple method eliminates the need for organic solvent extractions and nucleic acid precipitation, allowing for rapid purification of many samples simultaneously.

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Material Required

- Micro Pipettes Variable Volume 0.5-10µl, 10-100µl, and 100-1000µl
- Sterile pipette tips with aerosol barrier 2-20µl, 10-100µl, and 100-1000µl
- Disposable powder-free gloves
- Vortex mixer / Water bath
- Centrifuge with rotor for 1.5 ml reaction tubes
- 1.5ml/2ml centrifuge tubes
- 100% ethanol

Important Notes:

All purification steps should be carried out at room temperature.

All centrifugations should be carried out in a table-top micro-centrifuge at >12000 x g (12000-14000 rpm, depending on the rotor type).

Procedure:

Sample preparation

Brood stock eye stalk
Rinse the cut eye stalk with clean water. Put eye stalk into a 1.5ml tube that contains 0.6 ml tissue lysis buffer

Larvae, PL or Juvenile
Place about 10 -20 mg sample into a 1.5ml tube containing 0.6 ml Tissue lysis buffer.

Pleopod, pereopod, or gill of adult shrimp
Place 2 pieces into a 1.5ml tube containing 0.6 ml Tissue lysis buffer.

Tail or muscle of adult shrimp
Place a tail or about 20 mg muscle into a 2ml tube containing 0.6 ml Tissue Lysis buffer.

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1. Centrifuge at 8000rpm for 3 minutes.
2. Transfer 300µl of Supernatant into fresh 1.5ml centrifuge tube.
3. Add again 300µl of Binding buffer and vortex well. Incubate 2min at room temperature.
4. Add 600µl of [100%] ethanol and mix well by vortex for 30seconds. Spin down few seconds to bring down drops to bottom of the tube.
5. Pipette 600µl of sample into the PureFast® spin column. Centrifuge at 8000rpm for 1 min. Discard the flow-through and place the column back into the same collection tube.
6. Pipette remaining 600µl of sample into the PureFast® spin column. Centrifuge at 8000rpm for 1 min. Discard the collection tube containing filtrate. Place spin column into fresh collection tube.

7. Add 700µl of Wash buffer-1 [Ethanol added] to the PureFast® spin column. Centrifuge at 8000rpm for 1min and discard the flow-through. Discard the collection tube containing filtrate. Place spin column into fresh collection tube.

8. Add 700µl of Wash buffer-2 [Ethanol added] to the PureFast® spin column. Centrifuge at 8000rpm for 1min and discard the collection tube containing filtrate. Place spin column into fresh collection tube.

9. Repeat Wash buffer-2 wash once.

10. Centrifuge empty spin column attached with collection tube at **12000rpm** for an additional **1 min**. This step is essential to avoid residual ethanol. Discard collection tube.

11. Transfer the PureFast® spin column into a fresh 1.5 ml micro-centrifuge tube (not included).

12. Add following volume elution buffer as indicated below to the centre of PureFast® spin column membrane. Incubate 2 minute at room temperature.

    a) Post larvae = 400µl
    b) Brooder Eye stalk/tail/walking legs = 400µl
    c) Gills = 150µl
    d) adult animal muscle/Swing legs/Pancreas = 400µl

13. Centrifuge at 10000rpm for 1 min and discard the PureFast spin column. Centrifuge tube now contains the eluted nucleic acid. Either use the directly in PCR or store at -80°C for later analysis.

Recommendation for Real-time PCR:
Use 2.5 - 5µl of elute

Ordering information

Kit	Pack	Cat. No
HELINI™ PureFast Shrimp Nucleic acid Mini spin Prep kit [Shrimps]	25 Preps	2010
	50 Preps	2010
	100 Preps	2010

Manufactured and marketed by

HELINI *Biomolecules,*
Ohmlina, 26, 2nd Avenue,
Khuthubi Complex,
Vettuvankeni,
Chennai 600 115
www.helini.iin
helinibiomolecules@gmail.com

Technical doubts, please call +91-9382810333